

THE TOTAL SYNTHESIS OF THE ANTIBIOTIC MALONOMICIN (K16)

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(Received in UK 27 June 1977; Accepted for publication 15 July 1977)

Abstract—The total synthesis of the antibiotic malonomycin has been accomplished by a convergent synthesis in which the crucial step is a very mild coupling of the dipeptide sidechain to the 3-position of the pyrrolidin-2,4-dione ring.

Some years ago,¹ malonomycin (1) (formerly known as K16) was shown to be a representative of the continuously growing group of metabolites which all have in common a pyrrolidin-2,4-dione (tetramic acid) nucleus, acylated at the 3-position.

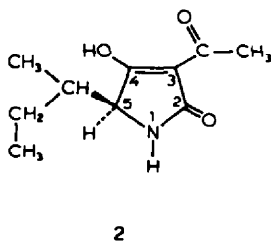
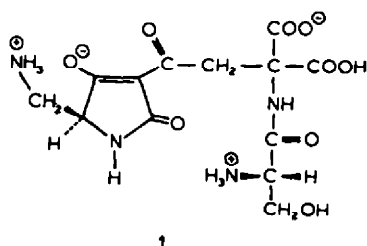
The first and still most simple member of this series is tenuazonic acid, the structure of which was determined as 3-acetyl-5-*sec*-butyl-4-hydroxy- Δ^3 -pyrrolin-2-one (2) by Stickings² (1959). In historical order, the other metabolites which have been shown to belong to this group are: streptolydigin (1963),³ erythroskyrin (1964),⁴ cyclopiiazonic acid (1968),⁵ tirandamycin (1971),⁶ ikarugamycin (1972),⁷ lipomycin (1973),⁸ olefinin (1973),⁹ magnesidin (1974),¹⁰ and very recently equisitine (1976).¹¹

of this moiety is still unknown but is the object of active investigation in this laboratory.

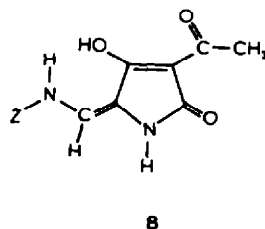
As far as we know up till now, only the total synthesis of tenuazonic acid¹⁵ and magnesidin,¹⁰ the more simple and stable representatives of this group of antibiotics, have been realized. Attempts to synthesize the tryptophane derived cyclopiiazonic acid¹⁶ are on the way. Not only as a keystone of the structure determination but also to explore the possibilities of synthesis of structural analogues, we undertook the total synthesis of malonomycin, expecting the construction of the sensitive aminomalonic acid part to constitute the main obstacle on the way.

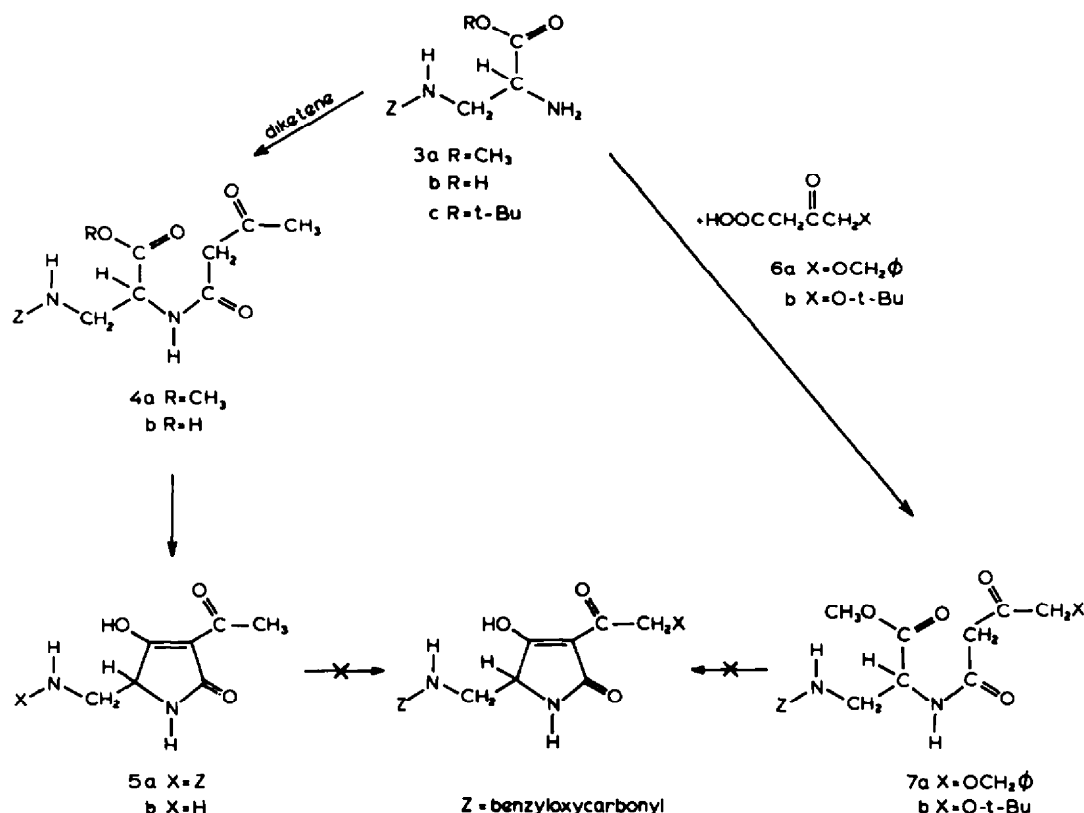
RESULTS AND DISCUSSION

At the outset, an obvious starting-point for the synthetic endeavour seemed to be the readily available 3-acetyl-5-*N*-benzyloxycarbonylaminoethyl-4-hydroxy- Δ^3 -pyrrolin-2-one (5a) (Scheme 1) which had served as an intermediate for the preparation of the valuable model compound 5b in the structure elucidation of malonomycin.¹ Proper functionalization of the acetyl group, followed by coupling with the anion of a suitable protected aminomalonic acid derivative, would then furnish a product which was expected to be easily transformed into a direct precursor of malonomycin. Furthermore, achievement of this goal would have provided considerable flexibility in attaching a variety of substituents leading to structural variations of malonomycin. However, all attempts to accomplish a selective transformation at the acetyl group of 5a were unsuccessful. E.g. bromination with *N*-bromosuccinimide, bromine-dioxane complex, pyridinium hydrobromide perbromide, pyrrolidone hydrotribromide, or elementary bromine, merely yielded starting material or intractable mixtures of products. Only with CuBr₂ in CHCl₃/EtOAc¹⁷ a single product was obtained in moderate yield, tentatively assigned structure 8 according to ¹H NMR and elemental analysis, most probably the result of bromination at C-5 followed by elimination of HBr.



The substituents at C-3 and C-5 of the common tetramic acid nucleus of these antibiotics are structurally very divergent and their biosynthetic origins largely unknown.¹² The metabolic source of the *sec*-Bu group of tenuazonic acid (2) together with C-4, C-5 and N-1 has been shown to be L-isoleucin.^{12a} Likewise, the results of preliminary biosynthetic investigations indicate that the aminomethyl group at C-5 of malonomycin has its origin in L-2,3-diaminopropanoic acid.¹³ A more conspicuous feature of the structure of 1 is the presence of the unique aminomalonic acid moiety,¹⁴ incorporated into the acyl substituent at C-3; this part of the molecule is of paramount importance for the biological activity of malonomycin as all activity is lost on decarboxylation.¹ The biosynthetic origin



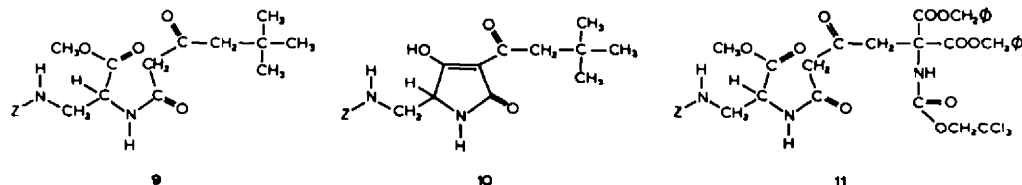


Scheme 1.

As the idea of a simply functionalized acetyltetramic acid derivative as a starting-point for the total synthesis was still appealing, we tried to cyclize 7a and 7b, the benzyloxy- and t-butoxy-substituted α -N-acetoacetyl derivatives of methyl *N*³-benzyloxycarbonyl-2,3-diaminopropanoate,¹⁸ prepared from 3a and benzyloxy- or t-butoxyacetoacetic acid, 6a and 6b, respectively (Scheme 1). However, neither under the original conditions of Lacey¹⁹ nor with other combinations of bases and solvents, this deceptively simple looking cyclization (analogous to the successful reaction 4a \rightarrow 5a) could be realized satisfactorily. Presumably, electronic effects of the alkoxy group are responsible for this failure; steric effects can be excluded, as cyclization of 9, prepared from 3a and γ -t-butylacetoacetic acid, was accomplished smoothly in the standard way,¹⁹ yielding 10 in good yield.

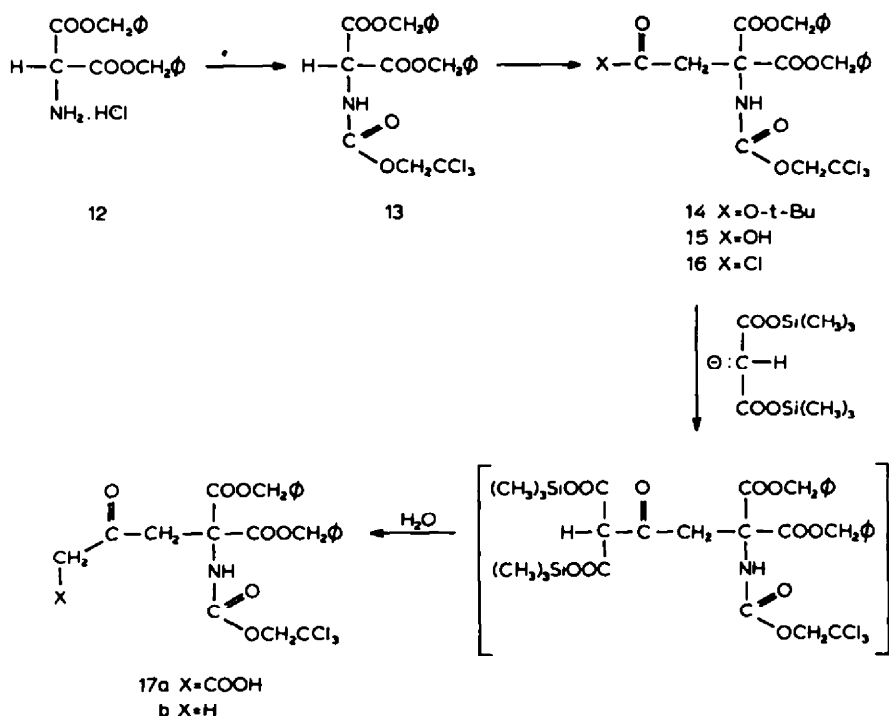
moiety is present, substituted with independently removable protecting groups for both the carboxy and amino functions.

The obvious approach to the synthesis of 11 seemed to be the coupling of 3a with the β -ketoacid 17a, which in its turn could be easily synthesized as shown in Scheme 2. Dibenzyl aminomalonate hydrochloride 12²⁰ was protected by means of trichloroethyl chloroformate²¹ in sodium bicarbonate solution, and alkylated with t-butyl chloroacetate in DMF to afford 14 in 78% overall yield. Boiling 14 for 4 min with a catalytic amount of *p*-toluenesulfonic acid in glacial acetic acid effected the removal of the t-Bu group. Treatment of the resulting carboxylic acid 15 with a tenfold excess of thionyl chloride in chloroform solution at room temperature furnished the acid chloride 16 in excellent yield.



The conversion of 9 to 10 illustrated that in principle the cyclization of derivatives of 4a carrying bulky substituents could be achieved, even, it was hoped, if provided with the appropriate functionality to elaborate the dipeptide side-chain. We therefore envisaged the synthesis of 11, in which, besides the diaminopropanoic and acetoacetic acid residues, also the aminomalononic acid

As conventional methods for synthesis of β -ketoacids, e.g. *via* (selective) hydrolysis of the corresponding alkyl β -keto-esters (Experimental, preparation of 6a and 6b) were expected to be troublesome in this particular case, a very mild and simple new method of general utility was developed, in which practically neutral conditions were preserved during the conversion of 16 into 17a. Thus, 16



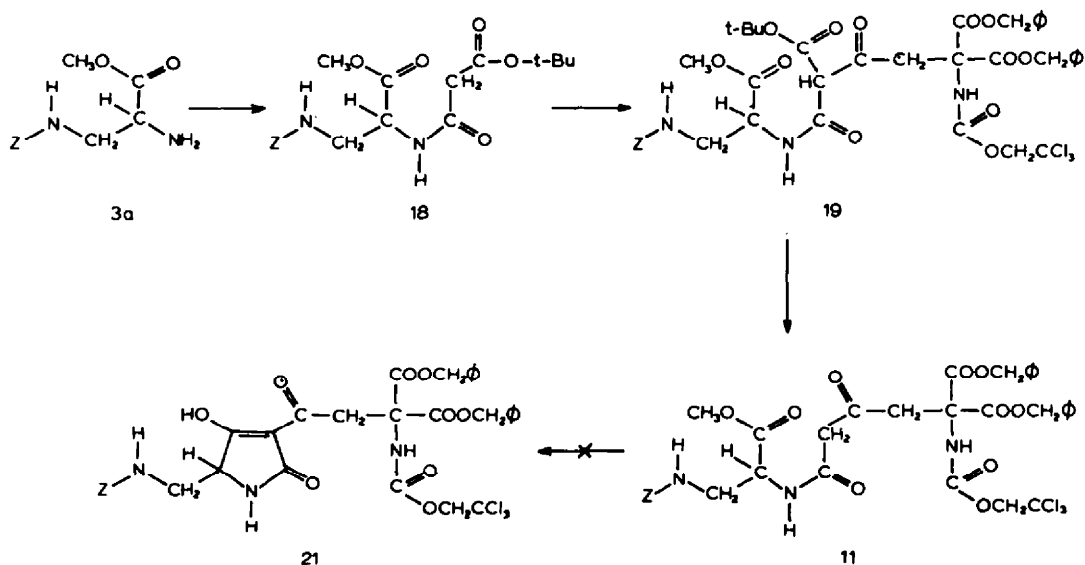
Scheme 2.

was quenched into a 100% excess of the mono-anion of bis(trimethylsilyl) malonate, followed by a short treatment with H_2O in which the intermediate triacyl compound was both hydrolyzed and decarboxylated to yield 17a in almost quantitative yield.²²

In numerous attempts with various coupling reagents, the condensation of 3a with 17a gave almost no yield of the desired 11, but instead the decarboxylation product 17b, the structure of which was proven by independent synthesis from 13 and bromoacetone. Successful coupling was only achieved *via* the benztriazolyl ester, formed *in situ* from 17a by the action of 1-hydroxybenztriazole (HOBT) and dicyclohexylcarbodiimide (DCC).²³

In a second approach, the synthesis of 11 could also be accomplished as shown in Scheme 3. Methyl *N*³-benzyloxycarbonyl-2,3-diaminopropanoate 3a was first coupled with *t*-butyl hydrogen malonate, followed by low temperature acylation with acid chloride 16 to give 19. Heating with *p*-toluenesulfonic acid in glacial acetic acid resulted in removal of the *t*-Bu group and simultaneous decarboxylation, yielding 11 in *ca.* 30% overall yield from 3a.

However, cyclization of 11 to 21 could not be effected. Neither under the standard conditions of Lacey¹⁹ (*cf.* conversion of 9 into 10) nor by employing other strong bases and solvents, such as potassium *t*-butoxide in



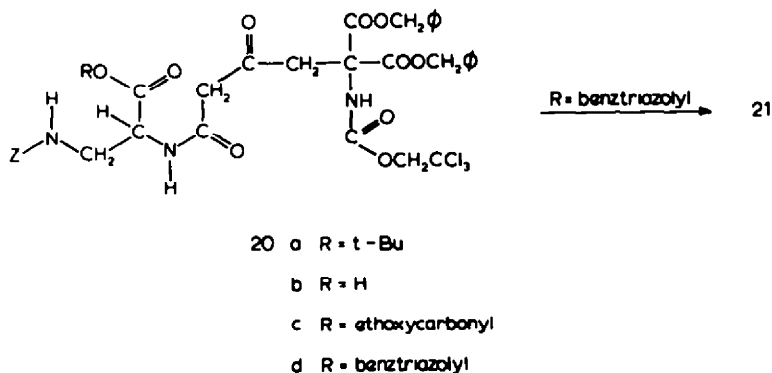
Scheme 3.

benzene or t-butanol, butyllithium in THF at low temperature, sodium hydride in DMF or benzene, or phase transfer agents and potassium carbonate in benzene/water system, any success was met. In most cases, not even a trace of starting material **11** could be recovered, probably due to the high base-sensitivity of the compounds involved. Obviously, base-induced decomposition of **11** and analogous compounds could only be prevented by application of a *weak* base; this, in turn, would presumably lead to a successful cyclization only if a very reactive ester instead of the methyl ester was present as the reaction partner.

Acid **20b** was considered to be a suitable starting material for the preparation of such an active ester. Its synthesis was realized by coupling t-butyl *N*³-benzyloxycarbonyl-2,3-diaminopropanoate **3c** with β -keto-acid **17**, followed by acid catalyzed removal of the protecting group. After some unsuccessful attempts, e.g. with mixed anhydrides (**20c**), the very reactive benzotriazolyl ester **20d**, prepared *in situ* from **20b**, HOBT and DCC in anhydrous THF at 0°, was treated with a slight excess of triethylamine. The UV spectrum

the crude reaction mixtures afforded only traces of malonomycin as indicated by TLC. An effective solution of this unexpected problem might have been found in the attachment of a seryl moiety to the aminomalonic acid part in an earlier stage of the synthesis, for instance by development of a β -ketoacid analogous to **17a**, but with a (protected) seryl residue instead of the amino-protecting trichloroethoxycarbonyl group of **20a**. Preliminary efforts in this direction were temporized when progress was achieved in a different approach to the total synthesis, i.e. the direct acylation of the tetramic acid nucleus.

Acyltetronic acids, the oxygen analogues of acyltetramic acids, have been made accessible in recent years by Bloomer *et al.* through acid-catalyzed acylation of the appropriate tetronic acids.²⁸ In contrast, synthetic work in the area of acyltetramic acids and derivatives still mostly follows lines described by Lacey as early as 1954, i.e. base-catalyzed cyclization of *N*-acetoacetyl-aminoester precursors.¹⁹ Only magnesidin has been prepared by TiCl₄-catalyzed acylation of a preformed pyr-



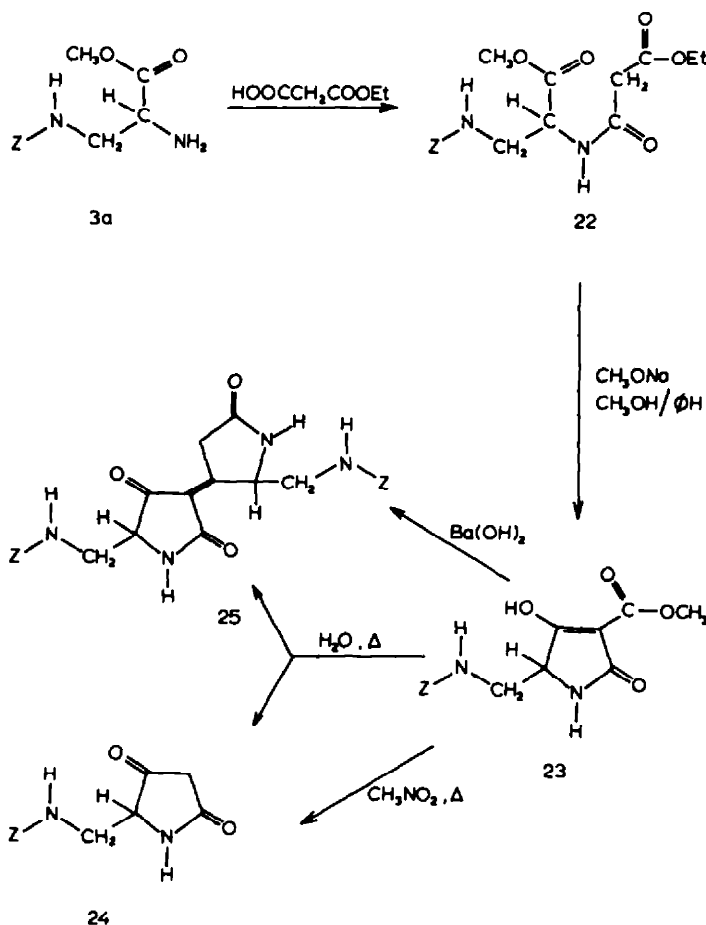
of the crude reaction mixture indicated the presence of the typical 3-acyltetramic acid chromophore (240 and 280 nm; the 240 nm band disappears on acidification^{1,3}). Working up was facilitated by the fact that the acidic cyclization product **21** (p*K*_A is expected to be *ca.* 3.5) was not extracted from ethyl acetate into sodium bicarbonate solution, so that most of the byproducts (HOBT, DCU) could be removed easily. Preparative TLC afforded **21** in a modest 18% yield, which presumably may be raised somewhat by proper choice of reaction conditions. Under the same conditions (but with 2 equivs of Et₃N) **4b**, prepared by direct condensation of **3b** and diketene,^{15,19,24} afforded **5a** in 41% yield.

The newly developed method for C-acylation is expected to be more generally applicable; in particular, it will be useful in those cases where, due to the presence of sensitive groups in the reactants, the usual acylation methods (e.g. those involving acid halides or anhydrides) fail (cf. also Scheme 7).

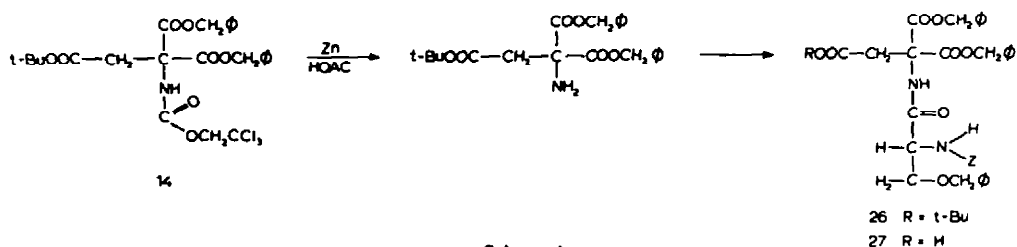
Removal of the protecting trichloroethoxycarbonyl group from **21** was easily accomplished with zinc in 90% acetic acid.²¹ However, coupling of the resulting amino ester with protected serine derivatives under a variety of conditions, including agents such as EEDQ,²⁵ DCC (with and without additives)²⁶ and DPPA (diphenylphosphoryl azide)²⁷ had little success; hydrogenolysis of

pyrrolidinedione nucleus with a simple straight-chain aliphatic acyl component.¹⁰ But as far as we are aware, no *base-induced* acylations of pyrrolidinediones have been carried out successfully, very probably due to a strong tendency of these compounds to dimerize under even mildly basic conditions. This became already evident in the preparation of **24** (Scheme 5), the requisite pyrrolidinedione precursor of malonomycin. Hydrolysis with simultaneous decarboxylation of **23** by Ba(OH)₂ according to the method of Ohta²⁹ led to self-condensation of the intermediate **24**, giving **25** almost exclusively. Careful hydrolysis of **23** by boiling in water for 6 min yielded **24** in moderate yield; prolonged heating lowered the yield considerably because of conversion to **25**. Highest yields of **24** (up to 90%) were obtained by heating **23** in nitromethane or acetonitrile containing traces of water.^{30,31}

As already mentioned, replacement of the trichloroethoxycarbonyl group in **21** by serine was not very successful. Therefore, it seemed advisable to perform the acylation of **24** with acid **27** or active derivatives thereof, in which serine is already present as an integral part. Acid **27** could be easily prepared (Scheme 6), starting from t-butyl ester **14**. Deprotection of **14** with zinc in 90% acetic acid, followed by coupling with *N*-benzyloxycarbonyl-*O*-benzyl serine³² afforded the oily t-butyl ester **26** (homogeneous



Scheme 5.



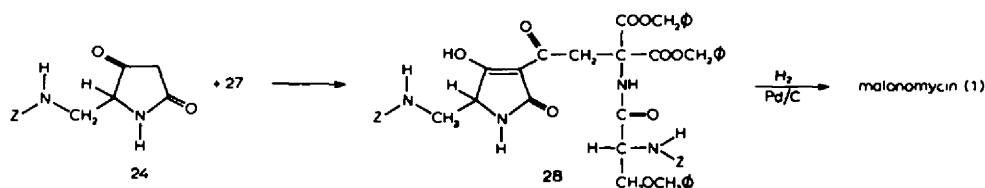
Scheme 6.

in TLC) which was treated with a catalytic amount of *p*-toluenesulfonic acid in boiling glacial acetic acid during 4 min to remove the *t*-Bu group. The resulting product was purified by preparative TLC to provide 27 in 45% overall yield from 14. As was to be feared, all attempts to accomplish an acylation of 24 at C-3 under normal, strongly basic conditions failed, mostly because of formation of dimer 25, but also because solubility problems prevented the use of solvents which were expected to stimulate C-acylation. Likewise, acylation under catalysis of Lewis acids such as TiCl_4 or BF_3 was unsuccessful.^{10,28}

At the same time, the cyclization of 20b to give acyltetramic acid 21 (Scheme 4) had demonstrated that active methylene compounds could be acylated under very mild conditions, i.e. in the presence of weak base (Et_3N) and at moderate temperature (25° or below) if benzotriazolyl esters were used as the acylating agent. Application of this principle to the particular problem of tetramic acid

acylation seemed promising because under these conditions dimerization of 24 to 25 was expected to be very slow. Thus, carboxylic acid 27 was converted *in situ* into the corresponding benzotriazolyl ester by treatment with HOBT and DCC in acetonitrile. Addition to the active methylene compound 24, followed by an equivalent of triethyl amine, afforded a reaction mixture from which after 48 hr at room temperature the C-acylated product 28 could be isolated by means of preparative TLC in ca. 40% yield (Scheme 7). Although the reaction product was pure by analytical TLC, the NMR spectrum was very difficult to interpret because of extensive line-broadening. The UV spectrum, showing the typical absorptions of the 3-acyltetramic acid chromophore, however, proved beyond all doubt that 28 indeed had been obtained.

As all protective groups of 28 are of the same type, i.e. benzyl and benzyloxycarbonyl groups, they can be removed in one step. After several attempts, this was



Scheme 7.

accomplished by hydrogenolysis at atmospheric pressure in methanol/water solution with 10% Pd on charcoal as a catalyst, activated by the presence of a little hydrochloric acid in order to remove even the stubbornly resistant benzyl group protecting the hydroxyl function of the seryl residue. The deprotection was performed at 0° to suppress decarboxylation as much as possible. The resulting product was purified by adsorption at pH 8.0 on a weakly basic anion-exchange column and elution with a gradient water/1N acetic acid. The active fractions were concentrated *in vacuo* leaving a white solid which according to IR, UV, ¹³C-NMR and chromatographic properties was indistinguishable from the natural antibiotic. The same was true of simple derivatives obtained by decarboxylation and/or dinitrophenylation.¹

The biological activity of the synthetic product, tested in mice infected with *Trypanosoma congolense*, was 25–50% of that of the microbiologically prepared product, owing to the fact that racemic derivatives of serine and 2,3-diaminopropanoic acid were used in the total synthesis. Due to the rather large margin of error of the biological test (mainly as a consequence of the availability of a too small quantity of synthetic material) it cannot be decided whether or not both asymmetric centers of malonomycin are required for full biological activity. As a matter of fact, it is not even certain whether the natural antibiotic is optically pure at the rather labile C-5 of the tetramic acid nucleus. However, the biological activity of the synthetic product must be considered as an extremely discriminative test of its structural identity, because even closely related derivatives such as the decarboxylation product of malonomycin lack this activity entirely.¹

Experiments leading to optically active precursors and therefrom to synthetic malonomycin with partial and total natural stereochemistry are on the way.

EXPERIMENTAL

All m.ps were determined on a Kofler hot stage apparatus under a Reichert microscope and are uncorrected. Unless otherwise stated, IR spectra were recorded in CHCl₃ with a Perkin-Elmer 237 spectrophotometer, UV spectra in 96% EtOH with a Perkin-Elmer 137 spectrophotometer, ¹H NMR spectra in CDCl₃ with a Varian A-60A spectrometer and ¹³C NMR spectra with a Varian XL-100 instrument (chemical shifts relative to TMS as an internal standard, δ = 0 ppm). All reactions were performed with purified reagents in a N₂ atmosphere.

t-Butyl N³ - benzyloxycarbonyl - 2,3 - diaminopropanoate (3c)

A soln of 3b¹⁸ (952 mg, 4 mmole), *t*-butyl acetate (60 ml) and perchloric acid (8.8 mmole) was kept at room temp. during 4 days.³³ After cooling in ice the mixture was extracted with 2N citric acid (4 × 5 ml) and the combined extracts immediately neutralized with solid NaHCO₃. Extraction with EtOAc, drying and evaporation *in vacuo* yielded 3c as a yellow oil (647 mg, 55%) which according to TLC (SiO₂; CHCl₃/acetone 3:1) was pure enough for conversion to 19; NMR: δ 1.42 (s, 9H), 1.95 (t, 3H), 3.0–3.6 (m, 3H), 5.03 (s, 2H), 6.05 (t, 1H) and 7.25 ppm (s, 5H).

Methyl N² - acetoacetyl - N³ - benzyloxycarbonyl - 2,3 - diaminopropanoate (4a)

A mixture of 3a, hydrochloride (3.77 g, 13.1 mmole), freshly distilled diketene (1.4 g, 25% excess) and NaHCO₃ (1.4 g) in 50% EtOH/H₂O (25 ml) was stirred for 2 hr at 0° and then another 2 hr at room temp.¹⁹ Extraction with CHCl₃ (3x), washing with brine, drying and evaporation *in vacuo*, yielded 4a as a colourless oil (4.53 g, 100%), ninhydrin-negative and pure by TLC (SiO₂; CHCl₃/acetone 4:1); NMR: δ 2.18 (s, 3H), 3.38 (s, 2H), 3.4–3.6 (m, 2H), 3.68 (s, 3H), 4.45–4.85 (m, 1H), 5.07 (s, 2H), 5.68 (t, 1H), 7.32 (s, 5H) and 7.61 ppm (d, 1H).

N² - Acetoacetyl - N³ - benzyloxycarbonyl - 2,3 - diaminopropanoic acid (4b)

To 3b¹⁸ (595 mg, 2.5 mmole) dissolved in such a quantity of 2N NaOH, that basic conditions were sustained during the reaction, freshly distilled diketene (270 mg, 30% excess) was added at 0° with vigorous stirring. After 2 hr at 0° and another 3 hr at room temp., the mixture was extracted with ether and the aqueous layer acidified with 2N HCl. After extraction with EtOAc and concentration *in vacuo*, 4b (810 mg) was obtained as a yellow oil (pure by TLC) which was used as such in the synthesis of 5a.

3 - Acetyl - 5 - N - benzyloxycarbonylaminoethyl - 4 - hydroxy - Δ³ - pyrrolin - 2 - one (5a)¹

(a) *By the method of Lacey.*¹⁹ A soln of 4a (772 mg, 2.3 mmole) in dry EtOH (1.4 ml) was added to Na (59 mg, 2.6 mmole) in dry EtOH (0.35 ml). After refluxing during 18 hr the mixture was cooled, poured on ice, and acidified with 2N HCl. The ppt was collected, washed with ice-water and dried over P₂O₅ *in vacuo*, yielding 5a (562 mg, 80%), m.p. 151–156°, almost pure by TLC (SiO₂; acetone/EtOH/4N NH₄OH 3:2:1). Crystallization from EtOH gave 460 mg of white crystals, m.p. 156–158°, identical by TLC, UV, IR and NMR with the material described below.

(b) *By the DCC/HOBT/Et₃N-method.* Crude 4b (810 mg, was dissolved in dry THF (10 ml) and then there were added at 0° sequentially HOBT (528 mg, 3.9 mmole), Et₃N (500 mg, 5 mmole) and DCC (575 mg, 2.8 mmole) in dry THF (2 ml). After standing for 1 hr at 0° and for 24 hr at room temp., the mixture was filtered and concentrated *in vacuo*. The residue was dissolved in NaHCO₃aq, washed with EtOAc, acidified to pH 1 and extracted again with EtOAc. After drying and evaporation, crystallization of the residue from MeOH furnished 5a (315 mg, 41%), m.p. 157–158.5°; positive FeCl₃-spot test; NMR (D₆-DMSO): δ 2.35 (s, 3H), 3.05–3.55 (m, 2H), 3.8–4.1 (m, 1H), 5.03 (s, 2H), 7.22 (t, 1H), 7.33 (s, 5H), 8.65 (s, 1H) and 11.17 ppm (s, 1H). (Found: C, 59.37; H, 5.32; N, 9.30. Calc. for C₁₅H₁₆N₂O₅: C, 59.20; H, 5.30; N, 9.21%).

3 - Acetyl - 5 - aminomethyl - 4 - hydroxy - Δ³ - pyrrolin - 2 - one (5b)¹

A partial suspension of 5a (2.55 g, 8.4 mmole) and 5% Pd-C (735 mg) in MeOH (70 ml) and glacial AcOH (1.75 ml) was stirred in a H₂ atmosphere, first at room temp. and then for 30 min at 50°. Cooling and filtration yielded a filter cake which was extracted with a small volume of 2N HCl. The filtered acidic soln was adjusted to pH 5 with Et₃N and the collected ppt of 5b dried over P₂O₅ *in vacuo* (1.1 g, 77%), m.p. > 300°; UV (max): 239 and 278 nm; NMR (D₂O + NaOD, pH 12): δ 2.33 (s, 3H), 2.89 (d, 2H, J = 4.2 cps) and 3.77 ppm (t, 1H, J = 4.2 cps); pK_a (H₂O): 1.93 (UV method).

4 - Benzyloxy - 3 - oxobutanoic acid (6a)

To a stirred suspension of NaH (24 g, 1 mole) in dry THF

(400 ml) was added dropwise (within 30 min at 10°) anhyd benzyl alcohol (250 ml), followed by a soln of ethyl 4-bromo-3-oxobutanoate³⁴ (41.8 g, 0.2 mole) in dry benzyl alcohol (100 ml). After stirring overnight at room temp. the mixture was neutralized with glacial AcOH (60 ml) under ice-cooling. Water was added and the mixture extracted with ether. The combined organic layers were washed with NaHCO₃ aq, dried, concentrated *in vacuo* and distilled to yield ethyl 4-benzyloxy-3-oxobutanoate (20.5 g, 43%), b.p. 120–125°/10⁻² mm; NMR: δ 1.20 (t, 3H), 3.48 (s, 2H), 4.10 (s, 2H), 4.13 (q, 2H), 4.55 (s, 2H) and 7.31 ppm (s, 5H).

This ethyl ester (11.8 g, 0.05 mole) was stirred with 1N NaOH (55 ml) and H₂O (85 ml) during 20 hr. The resulting soln was washed with ether (5 \times), cooled in ice and acidified with 1N H₂SO₄. Saturation with NaCl, extraction with ether (5 \times), washing with brine, drying and evaporation (temp. \leq 25°C) furnished **6a** (7.9 g, 76%) as a low-melting white solid, stable during months at -25°; IR (max): 3600–2400 (COOH), 1730 and 1710 cm⁻¹ (C=O); NMR: δ 3.49 (s, 2H), 4.11 (s, 2H), 4.52 (s, 2H) and 7.28 ppm (s, 5H).

4-t-Butoxy-3-oxobutanoic acid (**6b**)

This compound was prepared analogously to **6a** from ethyl 4-bromo-3-oxobutanoate³⁴ and *t*-BuOH via ethyl 4-*t*-butoxy-3-oxobutanoate (44% yield; b.p. 72–76°/1 mm; NMR: δ 1.21 (s, 9H), 1.28 (t, 3H), 3.54 (s, 2H), 4.02 (s, 2H) and 4.20 ppm (q, 2H), and saponification with NaOH (**6b**; 84% yield as a yellow oil; NMR: δ 1.22 (s, 9H), 3.60 (s, 2H), 4.07 (s, 2H) and 11.02 ppm (s, 1H)).

3-Acetyl-5-N-benzyloxycarbonylaminoethylidene-4-hydroxy- Δ^3 -pyrrolin-2-one (**8**)

Compound **5a** (365 mg, 1.2 mmole) and CuBr₂ (530 mg, 2.37 mmole) were suspended in CHCl₃ (3 ml) and EtOAc (6 ml), and boiled for 1 hr.¹⁷ After cooling to room temp. the ppt was removed by suction filtration and the filtrate evaporated to dryness *in vacuo*. The solid residue was treated with a small volume of acetone, leaving 52 mg of yellow crystals. The filter cake was extracted also with acetone and the extract evaporated to a small volume yielding another 70 mg of crystalline material, identical in all respects with the first crop. Total yield 122 mg of **8** (34%), m.p. 203–205°; UV (max): 281 and 340 nm; NMR (D₂O): δ 2.40 (s, 3H), 5.22 (s, 2H), 6.80 (d, 1H), 7.39 (s, 5H), 9.22 (s, 1H), 9.60 (d, 1H) and 10.56 ppm (s, 1H). (Found: C, 59.31; H, 4.60; N, 9.11. Calc. for C₁₅H₁₄N₂O₅: C, 59.60; H, 4.67; N, 9.27%).

Methyl N³-benzyloxycarbonyl-N²-(5,5-dimethyl-3-oxohexanoyl)-2,3-diaminopropanoate (**9**)

To a stirred soln of 5,5-dimethyl-3-oxohexanoic acid (395 mg, 2.5 mmole) in dry DMF (5 ml) was added a soln of **3a** (723 mg, 2.5 mmole) and Et₃N (250 mg, 2.5 mmole) in dry DMF (10 ml) under cooling in ice, followed by diphenylphosphoryl azide²⁷ (690 mg, 2.5 mmole). After stirring overnight the mixture was diluted with H₂O (150 ml) and extracted by EtOAc. Washing with 1N HCl, H₂O, NaHCO₃ aq and brine, drying and concentration *in vacuo* yielded **9** (830 mg, 85%) as a thick oil, reasonably pure by TLC (SiO₂; CHCl₃/ether 4:1) and NMR: δ 0.99 (s, 9H), 2.38 (s, 2H), 3.33 (s, 2H), 3.3–3.7 (m, 3H), 3.67 (s, 3H), 4.3–4.8 (m, 1H), 5.08 (s, 2H), 5.75 (t, 1H), 7.32 (s, 5H) and 7.65 ppm (d, 1H).

5-N-benzyloxycarbonylaminoethyl-3-(3,3-dimethylbutanoyl)-4-hydroxy- Δ^3 -pyrrolin-2-one (**10**)

A solution of **9** (240 mg, 0.61 mmole) in dry benzene (2 ml) was added to a stirred soln of NaH (55% in oil; 30 mg, 0.7 mmole) in dry MeOH (0.4 ml), and refluxed for 5 hr. Water was added under cooling and the aqueous phase extracted with ether. Acidification, extraction with EtOAc, drying and evaporation *in vacuo*, yielded **10** (174 mg, 73%). Recrystallization from benzene gave white crystals, m.p. 126–130°; UV (max): 242 and 283 nm; NMR (D₂O): δ 0.99 (s, 9H), 2.68 (s, 2H), 3.1–3.5 (m, 2H), 3.8–4.1 (m, 1H), 5.03 (s, 2H), 7.22 (t, 1H), 7.34 (s, 5H) and 8.5–9.2 ppm (m, 2H). (Found: C, 63.42; H, 6.70; N, 7.65. Calc. for C₁₉H₂₄N₂O₅: C, 63.32; H, 6.71; N, 7.79%).

Coupling product (**11**)

(a) *By coupling of 3a with 17a.* Compound **3a** (941 mg, 3.74 mmole; free amino-ester prepared from the hydrochloride by treatment with cold NaHCO₃ aq and extraction with EtOAc, drying and evaporation), HOBT (764 mg, 5.66 mmole) and **17a** (2.16 g, 3.74 mmole) were dissolved in dry THF (11 ml) at 0°. Then DCC (854 mg, 4.14 mmole) in dry THF (3 ml) was added with stirring.²³ After 1 hr at 0° and 4 hr at room temp., the precipitated dicyclohexylurea was filtered off and the filtrate concentrated *in vacuo*. The residue was dissolved in EtOAc and washed with NaHCO₃ aq, 2N citric acid, NaHCO₃ aq again, and brine. Drying and evaporation *in vacuo* gave 3.07 g of an oily product which was purified by preparative TLC (SiO₂; CHCl₃/ether 3:1) yielding **11** (1.72 g, 56%) and **17b** (399 mg, 20%), both fully identical with the independently synthesized products. (b) *By hydrolysis/decarboxylation of *t*-butyl ester 19.* Compound **19** (145 mg, 0.16 mmole) and *p*-toluenesulfonic acid (5 mg) in glacial AcOH (1 ml) were boiled for 4 min. Concentration *in vacuo*, redissolving in EtOAc, washing with NaHCO₃ aq, 0.5N HCl and brine, drying and evaporation *in vacuo* yielded **11** (117 mg, 90%) as a viscous oil, identical in all respects with the product obtained under (a); NMR: δ 3.32 (s, 2H), 3.4–3.65 (m, 2H), 3.70 (s, 3H), 3.80 (s, 2H), 4.45–4.85 (m, 1H), 4.61 (s, 2H), 5.0–5.2 (m, 3 \times 2H), 5.40 (t, 1H), 6.70 (s, 1H), and 7.1–7.5 ppm (m, 3 \times 5H + 1H).

Dibenzyl-N-trichloroethoxycarbonylamino malonate (**13**)

To **12²⁰** (2.68 g, 8 mmole) in sat. NaHCO₃ aq (22 ml) there was added dropwise in about 20 min trichloroethyl chloroformate^{21a} (1.91 g, 9 mmole) with vigorous stirring. After 3 hr the white ppt was collected, washed with water and dried *in vacuo* over P₂O₅ to yield **13** (3.71 g, 98%), m.p. 91–93° (from EtOH); NMR: δ 4.68 (s, 2H), 5.13 (d, 1H), 5.13 (s, 2 \times 2H), 6.28 (d, 1H) and 7.28 ppm (s, 2 \times 5H). (Found: C, 50.43; H, 3.94; Cl, 22.42; N, 2.82. Calc. for C₂₀H₁₈Cl₃NO₆: C, 50.60; H, 3.82; Cl, 22.40; N, 2.95%).

O¹-Benzyl-O⁴-*t*-butyl 2-benzyloxycarbonyl-N-trichloroethoxycarbonylaspartate (**14**)

Compound **13** (2.37 g, 5 mmole) dissolved in dry DMF (15 ml), was added to a stirred suspension of NaH (120 mg, 5 mmole) in dry DMF (25 ml). After 1.5 hr *t*-butyl chloroacetate (2.5 ml) was added and stirring continued for 18 hr. The mixture was then poured into dil. NaHCO₃ aq (200 ml) and the white ppt collected and dried over P₂O₅ *in vacuo*, yielding **14** (2.31 g, 79%), m.p. 123–124° (after recrystallization from EtOH); NMR: δ 1.36 (s, 9H), 3.43 (s, 2H), 3.64 (s, 2H), 5.14 (s, 2 \times 2H), 6.61 (s, 1H) and 7.27 ppm (s, 2 \times 5H). (Found: C, 52.95; H, 4.92; Cl, 18.13; N, 2.27. Calc. for C₂₂H₂₂Cl₃NO₆: C, 53.03; H, 4.79; Cl, 18.06; N, 2.38%).

O¹-Benzyl-O⁴-hydrogen 2-benzyloxycarbonyl-N-trichloroethoxycarbonylaspartate (**15**)

Compound **14** (1.175 g, 2 mmole) in glacial AcOH (12 ml) was boiled for 4 min with *p*-toluenesulfonic acid (60 mg). Concentration *in vacuo* and recrystallization from EtOH furnished **15** (900 mg, 85%), m.p. 142–144°; NMR: δ 3.58 (s, 2H), 4.62 (s, 2H), 5.13 (s, 2 \times 2H), 6.60 (s, 1H), 7.27 (s, 2 \times 5H) and 8.73 ppm (s, 1H). (Found: C, 49.69; H, 3.91; Cl, 20.10; N, 2.51. Calc. for C₂₂H₂₀Cl₃NO₆: C, 49.59; H, 3.78; Cl, 19.96; N, 2.63%).

Acid chloride **16**

A soln of **15** (532.5 mg, 1 mmole) and SOCl₂ (720 μ l, 10 mmole) in dry CHCl₃ (3 ml) was kept at room temp. for 4 days. Evaporation *in vacuo* left a colourless oil (550 mg, 100%) which solidified on standing, m.p. 78–86°. A sample for analysis was recrystallized from benzene/*n*-hexane, m.p. 86–88°; IR: 1797 cm⁻¹ (COCl); NMR: δ 4.15 (s, 2H), 4.62 (s, 2H), 5.12 (s, 2 \times 2H), 6.67 (s, 1H) and 7.28 ppm (s, 2 \times 5H).

β -Keto-acid **17a**

A soln of bis(trimethylsilyl) malonate²² (447 mg, 1.8 mmole) in dry ether (4 ml) at -60° was treated with *n*-BuLi (0.9 ml, 2N in hexane). After warming to 0°, **16** (497 mg, 0.9 mmole) in dry ether (2 ml) was added in one lot. After stirring for 10 min at 0° the

mixture was shaken with water and extracted with ether. Drying and evaporation yielded a yellow oil, which was taken up in a small volume of CHCl_3 and diluted with hexane to give 17 (480 mg, 92%) as white crystals, m.p. 105–115° (dec). An analytical sample was prepared by repeated crystallization from CHCl_3 /hexane, m.p. 114–117°; NMR: δ 3.43 (s, 2H), 3.87 (s, 2H), 4.62 (s, 2H), 5.15 (s, 2×2H), 6.68 (s, 1H), and 7.30 ppm (s, 2×5H). (Found: C, 49.82; H, 4.09; Cl, 19.05; N, 2.30. Calc. for $\text{C}_{24}\text{H}_{22}\text{Cl}_3\text{NO}_5$: C, 50.15; H, 3.86; Cl, 18.51; N, 2.44%).

Acetonyl compound 17b

A soln of 13 (1.19 g, 2.5 mmole) in dry DMF (10 ml) was added to a stirred suspension of NaH (62 mg, 2.6 mmole) in dry DMF (5 ml). When the H_2 evolution had ceased (ca. 1 hr) bromoacetone (514 mg, 50% excess) was added and stirring continued overnight. The clear mixture was diluted with NaHCO_3 aq (100 ml) and extracted with ether. Washing with brine, drying and evaporation *in vacuo* gave 1.28 g of a yellow oil which solidified on standing. Recrystallization from EtOH yielded 17b (1.08 g, 80%), m.p. 75–78°; NMR: δ 1.99 (s, 3H), 3.70 (s, 2H), 4.63 (s, 2H), 5.11 (s, 2×2H), 6.74 (s, 1H) and 7.25 ppm (s, 2×5H). (Found: C, 51.87; H, 4.16; Cl, 20.18; N, 2.50. Calc. for $\text{C}_{23}\text{H}_{22}\text{Cl}_3\text{NO}_7$: C, 52.05; H, 4.18; Cl, 20.04; N, 2.64%).

Methyl N^3 - benzyloxycarbonyl - N^2 - (O^3 - *t* - butyl malonyl) - 2,3 - diaminopropanoate (18)

t-Butyl hydrogen malonate (544 mg, 3.4 mmole) was added to a stirred soln of 3a (free aminoester; 852 mg, 3.4 mmole) and EEDQ²⁵ (900 mg, 3.6 mmole) in dry THF (25 ml). After 20 hr at room temp. the mixture was concentrated *in vacuo* and the residue dissolved in ether, washed with ice-cold 2N citric acid, H_2O , NaHCO_3 aq and brine, respectively. Drying and evaporation *in vacuo* gave 18 (1.12 g, 84%) as a thick oil, homogeneous in TLC; NMR: δ 1.43 (s, 9H), 3.20 (s, 2H), 3.4–3.7 (m, 2H), 3.65 (s, 3H), 4.4–4.85 (m, 1H), 5.07 (s, 2H), 5.85 (t, 1H), 7.32 (s, 5H) and 7.77 ppm (d, 1H).

Acylation product 19

To a stirred soln of 18 (392 mg, 1 mmole) in dry ether (2 ml) at –60°, BuLi (0.5 ml of a 2N soln in hexane) was added. After 15 min, 16 (552 mg, 1 mmole) in dry ether (1 ml) was quickly added at 0° and stirring continued for 1 hr at 0° and 1 hr at room temp. Acidification with 1N HOAc (4 ml), extraction with ether, washing with NaHCO_3 aq and brine, drying and evaporation *in vacuo* yielded a glassy product (840 mg) which was purified by TLC (SiO_2 ; CHCl_3) to furnish 19 (362 mg, 40%) as a colourless glass; NMR: δ 1.50 (s, 9H), 3.4–3.65 (m, 2H), 3.72 (s, 3H), 4.15 (s, 2H), 4.45–4.85 (m, 1H), 4.63 (s, 2H), 5.0–5.2 (m, 3×2H), 5.25 (t, 1H), 6.63 (s, 1H), 7.15–7.5 (m, 3×5H + 1H) and 9.75 ppm (m, ca. 1H).

Coupling product 20a

This product was prepared from 3c and 17a analogous to the synthesis of 11 (reaction time 20 hr at room temp.), yield (after preparative TLC) 64%; NMR: δ 1.43 (s, 9H), 3.34 (s, 2H), 3.4–3.8 (m, 2H), 3.85 (s, 2H), 4.3–4.8 (m, 1H), 4.63 (s, 2H), 5.13 (m, 3×2H), 5.70 (t, 1H), 6.79 (s, 1H) and 7.1–7.5 ppm (m, 3×5H + 1H).

Acid 20b

This product was prepared from 20a analogous to the synthesis of 11 (method b), yield 83%, homogeneous in TLC (SiO_2 ; EtOAc/EtOH 4:1); IR: 3700–2700 cm^{-1} (COOH); NMR: no *t*-Bu group.

Cyclization product 21

A stirred soln of 20b (769 mg, 0.96 mmole) in dry THF (4 ml) was treated at 0° with HOBT (185 mg, 1.42 mmole) and DCC (220 mg, 1.1 mmole). After 1 hr at 0° and 2 hr at room temp., Et_3N (100 mg, 1 mmole) was added and the mixture was stirred during 2 days. After filtration and dilution with EtOAc, the mixture was washed with NaHCO_3 aq, H_2O , 1N citric acid and brine, respectively. Drying and concentration *in vacuo* gave 730 mg of a viscous oil which was purified by TLC (SiO_2 ;

CHCl_3 /acetone 1:1), yielding 21 (131 mg, 18.4%) as an amorphous powder (dec. at 130–135°); positive FeCl_3 -spot test; UV (max): 240 (ϵ 9600) and 280 nm (11.250).

Methyl N^3 - benzyloxycarbonyl - N^2 - (O^3 - ethyl malonyl) - 2,3 - diaminopropanoate (22)

To 3a (2.21 g, 7.7 mmole) and Et_3N (774 mg, 7.7 mmole) in CH_2Cl_2 (15 ml) was added at 0° a soln of ethyl hydrogen malonate (1.01 g, 7.6 mmole) in CH_2Cl_2 (6 ml) and then EEDQ (2.02 g, 8.1 mmole). After stirring for 6 hr the mixture was washed with 1N HCl, NaHCO_3 aq and H_2O . Upon concentration *in vacuo*, 22 (2.49 g, 89%) was obtained as an oil, almost pure according to TLC and NMR: δ 1.23 (t, 3H), 3.28 (s, 2H), 3.68 (s, 3H), 3.45–3.75 (m, 2H), 4.15 (q, 2H), 4.45–4.85 (m, 1H), 5.07 (s, 2H), 5.68 (t, 1H), 7.30 (s, 5H) and 7.73 ppm (d, 1H).

5 - N - Benzyloxycarbonylaminomethyl - 4 - hydroxy - 3 - methoxycarbonyl - Δ^3 - pyrrolin - 2 - one (23)

A soln of 22 (8.82 g, 0.024 mole) and NaH (1.22 g of a 57% suspension in oil, 0.03 mole) in dry MeOH (16 ml) and dry benzene (40 ml) was stirred and boiled for 3 hr. After cooling, H_2O (50 ml) was added and the organic layer separated. The aqueous layer was acidified to pH 1 and the ppt collected and recrystallized from MeOH to yield 23 (4.36 g, 57%), m.p. 145–148°; positive FeCl_3 -spot test; NMR (D_2 -DMSO): δ 2.9–3.85 (m, 2H), 3.67 (s, 3H), 3.85–4.20 (m, 1H), 5.06 (s, 2H), 7.22 (t, 1H), 7.33 (s, 5H) and 10.08 ppm (s, 1H). (Found: C, 56.15; H, 5.28; N, 8.91. Calc. for $\text{C}_{15}\text{H}_{16}\text{N}_2\text{O}_6$: C, 56.24; H, 5.03; N, 8.75%).

5 - N - Benzyloxycarbonylaminomethylpyrrolidin - 2,4 - dione (24)

A soln of 23 (1.63 g, 5.1 mmole) in nitromethane (32 ml) containing H_2O (180 μl , 10 mmole) was boiled for 30 min. The residue obtained after evaporation of the solvent *in vacuo* was recrystallized from CHCl_3 , yielding 24 (1.17 g, 88%), m.p. 131–135°; NMR (CD_3CN): δ 2.88 (2×s, 2H), 3.25–3.5 (m, 2H), 3.8–4.25 (m, 1H), 5.07 (s, 2H), 5.83 (t, 1H), 6.90 (d, 1H) and 7.33 ppm (s, 5H). (Found: C, 59.39; H, 5.49; N, 10.63. Calc. for $\text{C}_{15}\text{H}_{14}\text{N}_2\text{O}_4$: C, 59.54; H, 5.38; N, 10.68%).

O^1 - Benzyl O^4 - *t* - butyl N - (O^3 - benzyl - N - benzyloxycarbonylseryl) - 2 - benzyloxycarbonylaspartate (26)

To a stirred soln of 14 (1.2 g, 2.04 mmole) in 90% AcOH (60 ml) there were added over a period of 15 min three portions of 1.5 g each of Zn powder at 0°. After stirring for 1 hr at 0° and for 1.5 hr at room temp., the mixture was filtered, the filtrate concentrated *in vacuo* and the residue dissolved in EtOAc and H_2O . The organic layer was washed with 1N NH_4OH and evaporated *in vacuo* to dryness, yielding the deprotected product O^1 -benzyl O^4 - *t* - butyl 2 - benzyloxycarbonylaspartate (843 mg, 100%); (NMR: δ 1.38 (s, 9H), 2.55 (s, 2H), 3.09 (s, 2H), 5.18 (s, 2×2H) and 7.34 ppm (s, 2×5H)). This compound (843 mg, 2.04 mmole) and N-benzyloxycarbonyl - O - benzylserine³² (671 mg, 2.04 mmole) were dissolved in dry THF (9 ml) and a soln of DCC (430 mg, 2.09 mmole) in dry THF (1 ml) was added at about –10°. After stirring for 1 hr at –10°, the mixture was kept for 2 days at 0°. Filtration and concentration *in vacuo* gave an oil which was redissolved in EtOAc and washed with cold 1N HCl and sat. NaHCO_3 aq. Drying and evaporation *in vacuo* yielded 26 (1.51 g, 100%) as a viscous oil, almost pure by TLC (SiO_2 ; $\text{CHCl}_3/\text{CCl}_4$ 5:2); NMR: δ 1.32 (s, 9H), 3.48 (s, 2H), 3.4–3.9 (m, 2H), 4.45 (s, 2H), 4.1–4.6 (m, 1H), 5.10 (m, 3×2H), 5.64 (d, 1H), 7.1–7.4 (m, 4×5H) and 8.17 ppm (s, 1H).

O^1 - Benzyl O^4 - hydrogen N - (O^3 - benzyl - N - benzyloxycarbonylseryl) - 2 - benzyloxycarbonylaspartate (27)

This compound was synthesized from 26 analogous to the preparation of 11 (method b), yield 93%, purification (with substantial loss) by preparative TLC (SiO_2 ; CHCl_3 /acetone 4:1); NMR: δ 3.2–3.9 (m, 4H), 4.15–4.6 (m, 1H), 4.37 (s, 2H), 5.08 (m, 3×2H), 5.87 (d, 1H), 7.1–7.4 (m, 4×5H), 8.18 (s, 1H) and 9.82 ppm (s, 1H).

3 - [O¹ - Benzyl N - (O¹ - benzyl - N - benzyloxycarbonylseryl) - 2 - benzyloxycarbonyl - 4 - aspartyl] - 5 - N - benzyloxycarbonyl-aminomethyl - 4 - hydroxy - Δ^3 - pyrrolin - 2 - one (28)

To 27 (890 mg, 1.33 mmole) and HOBT (196 mg, 1.45 mmole) in dry acetonitrile (30 ml) was added at 0° DCC (299 mg, 1.45 mmole) in dry acetonitrile (15 ml). After stirring for 45 min at 0° and for 1 hr at room temp., the mixture was added to a soln of 24 (349 mg, 1.33 mmole) and Et₃N (145 mg, 1.45 mmole) in dry acetonitrile (225 ml). After 3 days at room temp., the mixture was filtered and the filtrate concentrated *in vacuo* to dryness. The residue was dissolved in EtOAc, washed with NaHCO₃ aq, 2N citric acid and H₂O, sequentially, dried and concentrated *in vacuo* yielding 1.07 g of a viscous oil. Further purification by preparative TLC (SiO₂; CHCl₃/EtOH/acetone/25% NH₄OH 9:1:1:0.1) gave 28 (486 mg, 40%); UV (max)(CH₃OH): 243 and 284 nm; positive FeCl₃-spot test.

5 - Aminomethyl - 3 - (2 - carboxy - N - seryl - 4 - aspartyl) - 4 - hydroxy - Δ^3 - pyrrolin - 2 - one (malonomycin) (1)

A soln of purified 28 (138 mg, 0.15 mmole) in MeOH (3.5 ml) containing a few drops of 6N HCl was vigorously stirred for 18 hr at 0° with 10% Pd-C (25 mg) in a H₂ atmosphere. After filtration and concentration *in vacuo*, the residue was dissolved in 10 mM NH₄OAc-soln (100 ml) and adsorbed on DEAE-Sephadex A-25 (column of 35 × 1.5 cm; adsorbents swollen in 0.5 M NH₄OAc and equilibrated with 10 mM NH₄OAc-soln). After washing with H₂O (pH 7), the product was eluted with a linear gradient H₂O/1N HOAc (total of 600 ml; flow rate 30 ml/hr. The fractions containing the product were collected and evaporated *in vacuo* at 25°, yielding racemic 1 (48 mg, 86%) (contaminated with a very small amount of its decarboxylation product), fully identical with the natural product¹ by TLC (SiO₂; EtOH/1N NH₄OH 3:2), UV, IR and ¹³C-NMR (D₂O + NaOD, pD 8.5): δ 40.4, 44.5, 55.6, 56.6, 62.3, 67.6, 102.9, 169.2, 175.3, 178.5, 193.5 and 194.4 ppm.

Acknowledgement—We thank Gist-Brocades N.V., Delft, for generous gifts of the antibiotic, and for measurement and comparison of the biological activities of synthetic and natural malonomycin.

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